

# Assays for gene expression and protein production

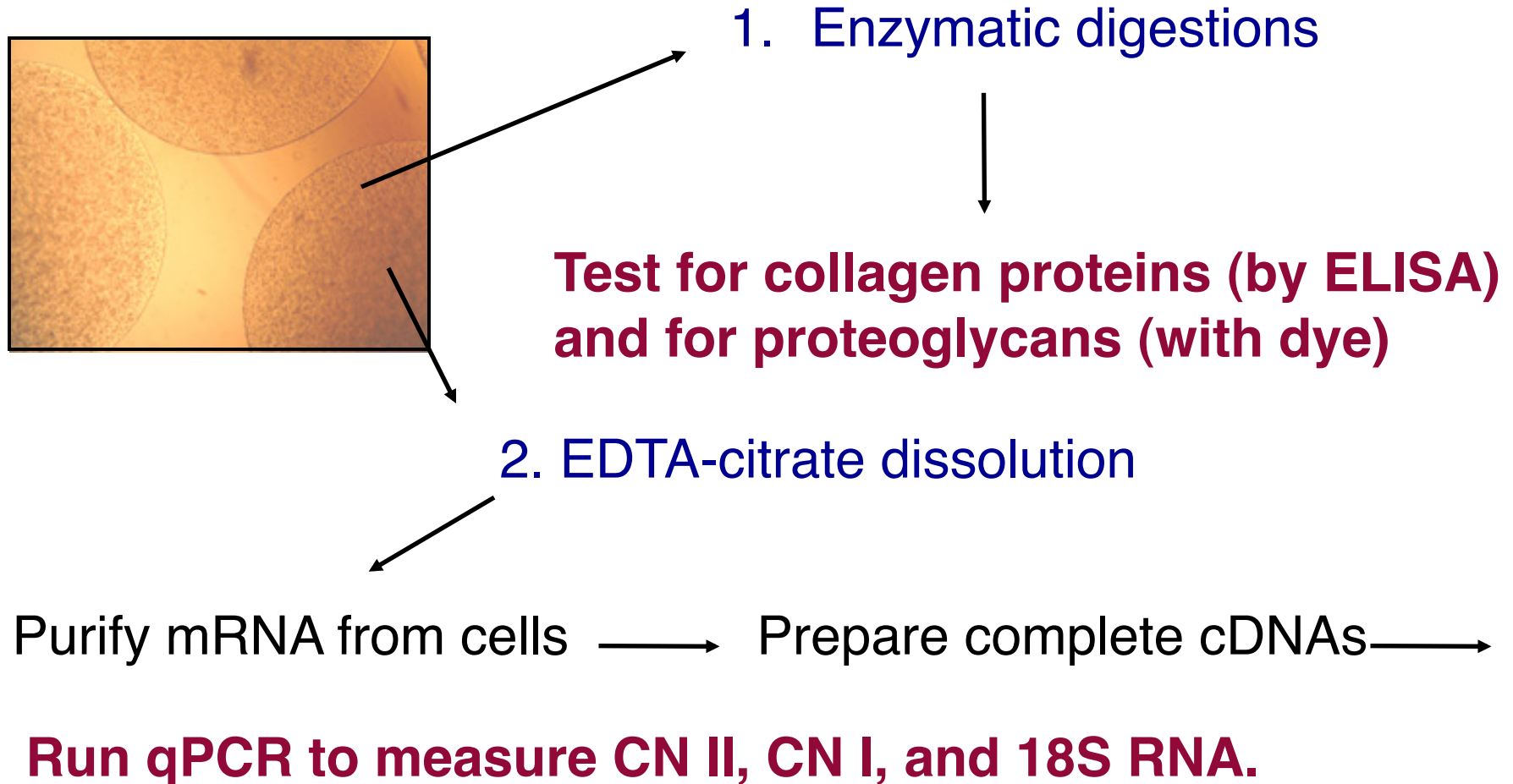
Module 3, Lecture 5

20.109 Spring 2011

# Topics for Lecture 5

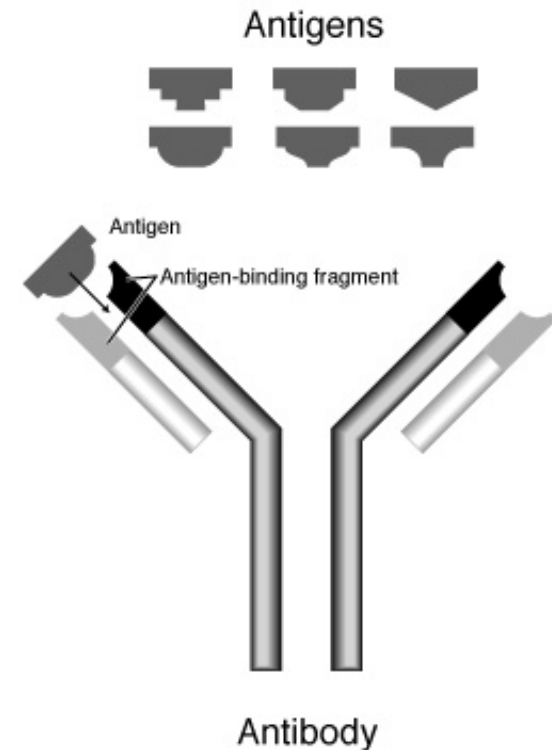
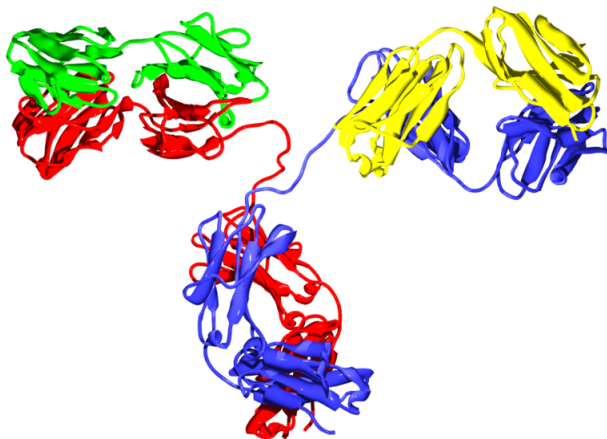
- Measuring protein levels
- Measuring transcript levels
- Imaging assays

# Module overview: 2<sup>nd</sup> half



# Antibodies are specific and diverse

- Specificity
  - variable region binding,  $K_D \sim \text{nM}$
  - linear or conformational antigens
- Diversity
  - gene recombination
- Production
  - inject animal with antigen, collect blood
  - hybridomas (B cell + immortal cell)



Public domain images  
(Wikimedia commons)

# Day 5-7: protein analysis by ELISA

- ELISA: enzyme-linked immunosorbent assay

- specific
- sensitive
- multiple kinds

“blocking” step  
also needed

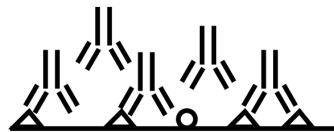
△ = protein  
of interest

## Indirect ELISA

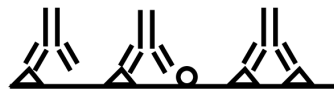
bind antigen



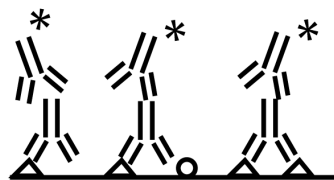
add 1° antibody



wash



add 2° antibody, wash



## Sandwich ELISA

bind capture antibody



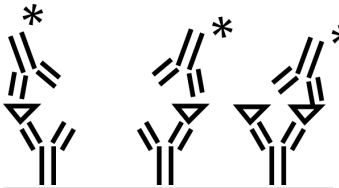
add antigen



wash

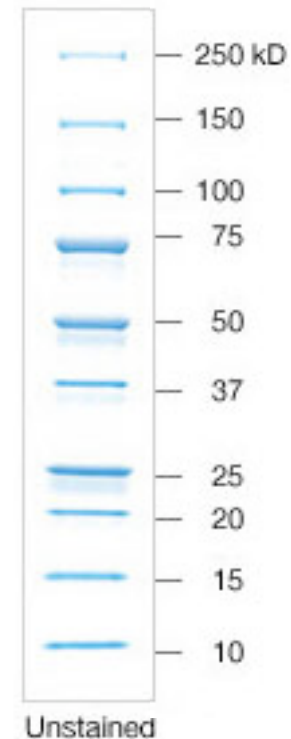


add detection antibody, wash



# Protein gels: SDS-PAGE

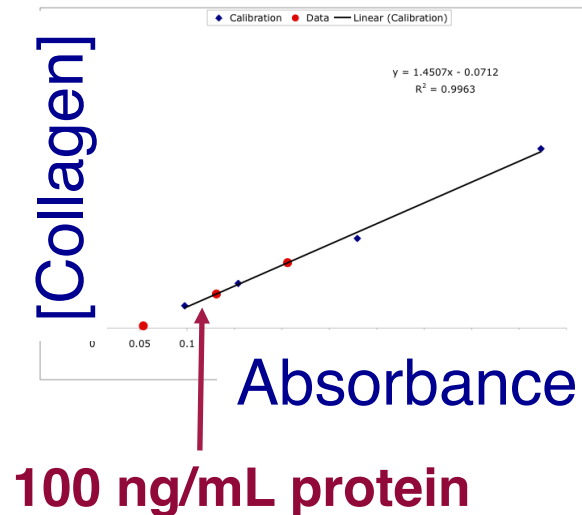
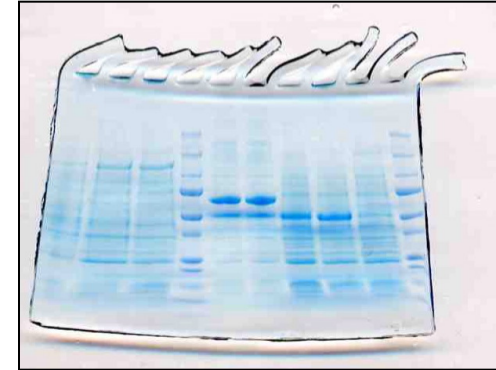
- Polyacrylamide gel electrophoresis
  - separates proteins
  - by size, shape, charge
- Sample preparation
  - SDS to coat with negative charge
  - $\beta$ -Me to break disulfide bonds
  - boiling to further denature
- Visualization: Coomassie stain
  - binds certain AA



protein ladder,  
bio-rad.com

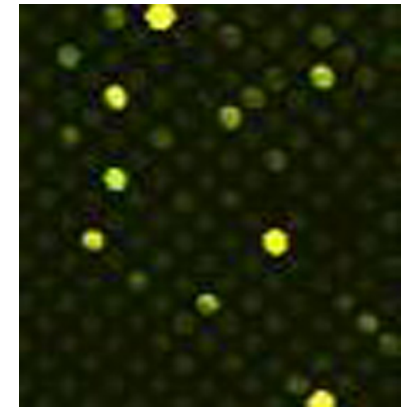
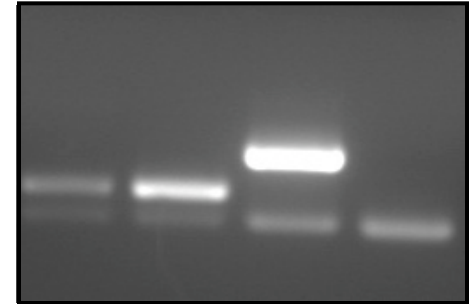
# Common protein-level assays

- PAGE
  - simple and low cost
  - Coomassie detection limit  $\sim 0.3$ -1  $\mu\text{g}/\text{band}$  (2-5  $\text{ng}/\text{band}$  for silver staining)
  - cannot distinguish two proteins of same MW
- Western blot
  - identifies specific protein
  - detection limit  $\sim 1$  pg (chemiluminescent)
  - only simple for denatured proteins
- ELISA
  - detects native state proteins
  - quantitative
  - high throughput



# Common transcript-level assays

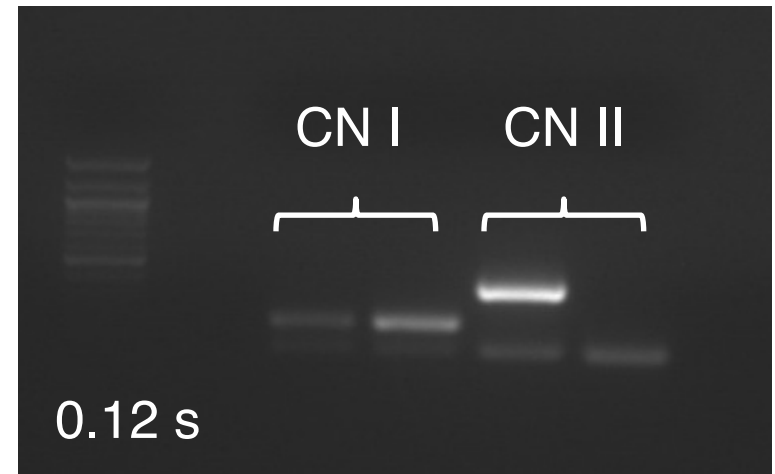
- RT-PCR (end-point)
  - simple, low cost
  - can be semi-quantitative
- Microarrays (end-point)
  - high cost, need specialty equipment
  - complicated and fraught analysis
  - high throughput
- q-PCR (real-time)
  - some special equipment, medium cost
  - highly quantitative
  - multiplexing potential
  - requires optimization (primers)





# End-point RT-PCR

- Co-amplification in one tube
  - Collagen + GAPDH
- Optimize primers
  - no cross-hybridization
  - similar signals (vary [primer])
  - similar efficiency
- Reliability issues
  - must be in exponential phase
  - sensitive to change in [RNA]
- Visualize on a gel
  - measure band intensity/area
  - low dynamic range

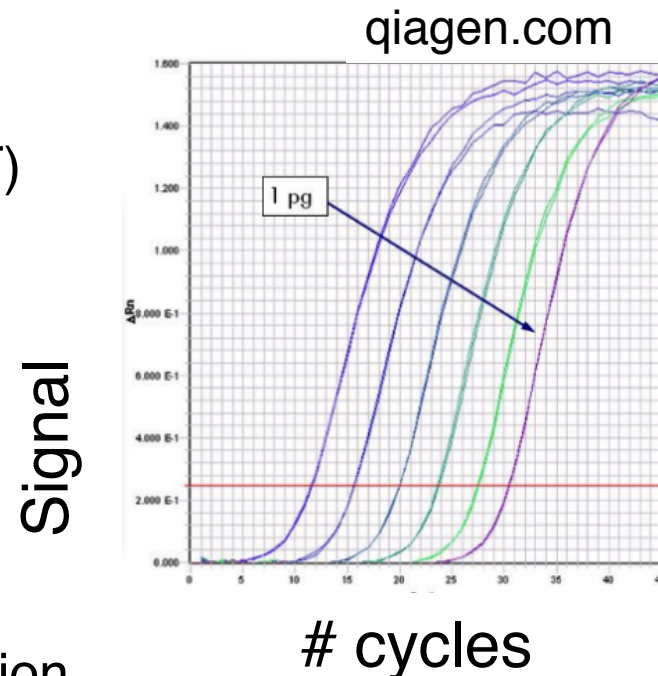


Collagen (upper band)  
GAPDH (lower band)

*Which sample is from chondrocytes, and which from stem cells?*

# Introduction to qPCR

- Real-time tracking of [DNA]
- Uses probes that fluoresce
  - when bind to any DNA
  - when bind to specific DNA (FRET)
- How and why does [DNA] change during PCR?
  - first plateau
  - exponential phase
  - second plateau
  - detection limit
  - competition, reagent limits, inhibition



- Starting point for analysis: threshold cycle  $C_T$

*Current Protocols in Cell Biology, Molecular Biology*

# Interlude: intersection of science and commerce

## Patenting genes

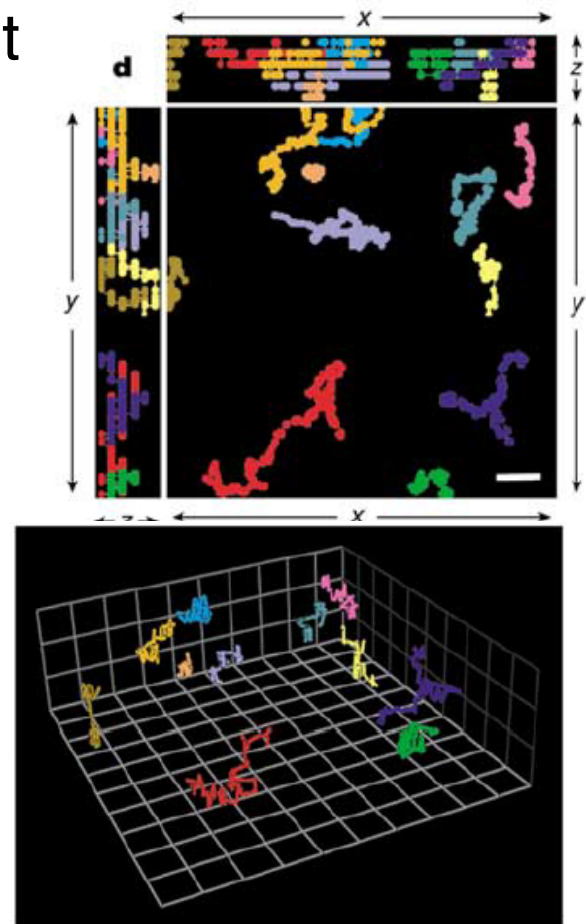
“Judge invalidates human gene patent”  
*NY Times* March 2010

“Metastasizing patent claims on BRCA1”  
*Genomics* May 2010

# Day 5-6: image analysis

- Imaging data is often high throughput
  - 4D: time,  $x$ - $y$ - $z$
  - requires computation, *and*
  - human design/interpretation
- Many available analysis packages
  - some ~ \$20-30K
  - NIH ImageJ = free
- Your analyses
  - automated cell counts
  - optional: explore other features

Images from: T.R. Mempel, et al. *Nature* **427**:154 (2004)



# Fluorescence microscopy

- Light source
  - Epifluorescence: lamp (Hg, Xe)
  - Confocal: laser (Ar, HeNe)
  - 2-photon: pulsed laser
- Filter cube
  - Excitation
  - Dichroic mirror
  - Emission
  - Band-pass vs. long-pass
- Detection
  - CCD camera

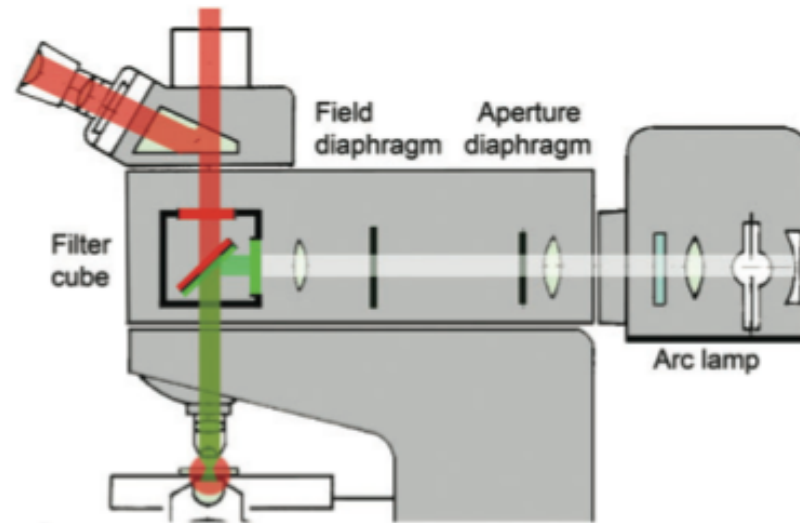
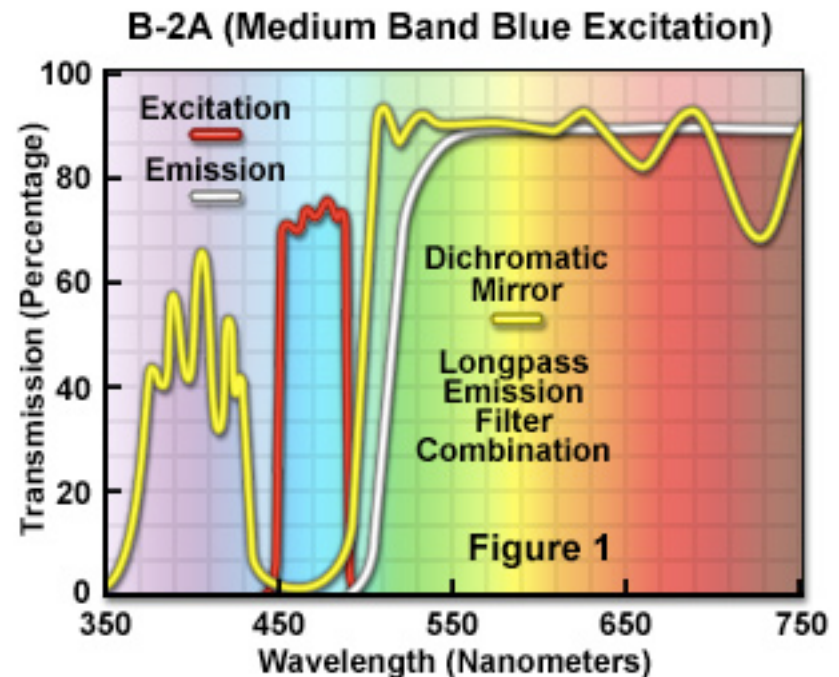
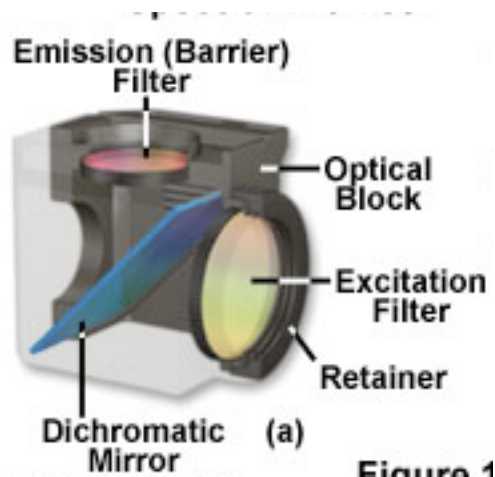


Image from: Lichtman & Conchello, *Nature Methods* 2:910 (2005)

# Specifications for Day 3 imaging

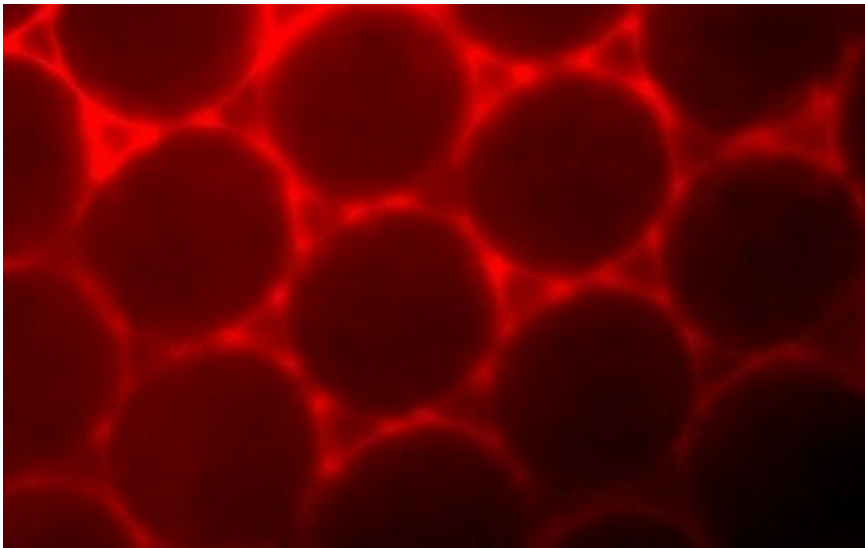
- Live/Dead Dyes
  - Green 490 ex, 520 em
  - Red 490 ex, 620 em
- Excitation 450-490 nm
- Dichroic 500 nm
- Emission 515<sup>+</sup> nm



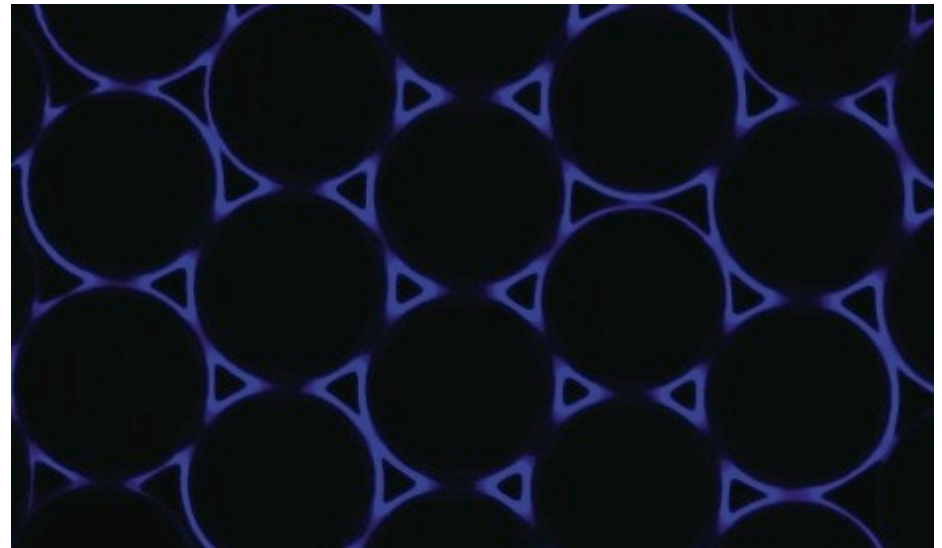
Images from: Nikon microscopy  
website: [www.microscopyu.com](http://www.microscopyu.com)

# Types of microscopy

- Epifluorescence: noisy due to out-of-plane light
- Confocal: pinhole rids out-of-plane light
- 2-photon: femtoliter volume excited; good depth (IR)

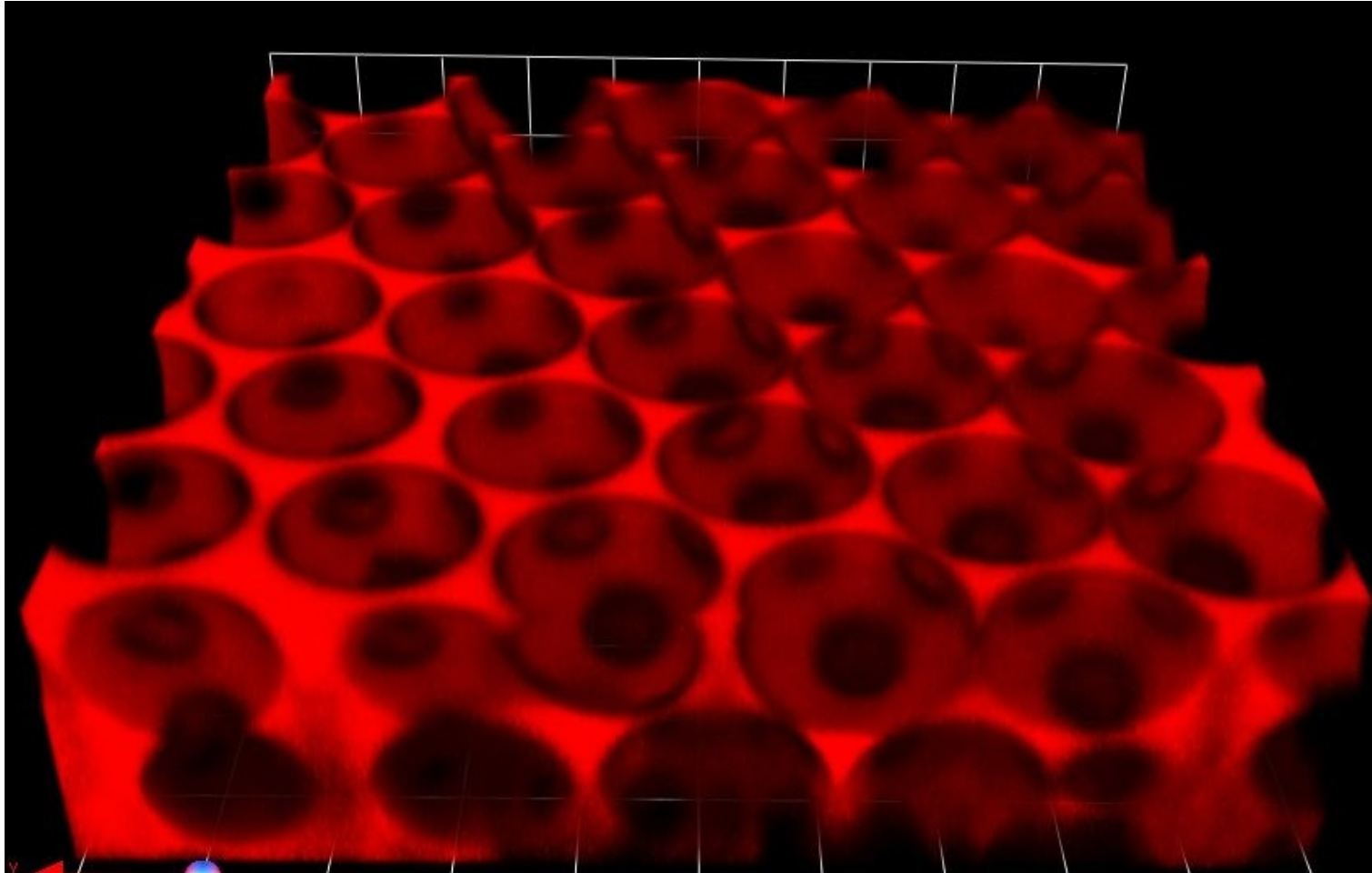


Epifluorescence



Confocal

# Confocal uscopy permits 3D reconstruction





# Lecture 5: conclusions

- Antibodies to diverse targets (e.g., proteins) can be made and used for detection/measurement.
- Trade-offs exist (e.g., between simplicity and accuracy) for different transcript-level assays.
- Fluorescence imaging is a powerful tool for studying cells and materials.

Next time: cartilage TE, from *in vitro* and *in vivo* models to the clinic; qPCR analysis.